

Lot No.: **34M**

Lot-specific information

[www.olerup-ssp.com](http://www.olerup-ssp.com)

## **Olerup SSP<sup>®</sup> DQA1\*02,05;DQB1\*02,03:02**

Product number: 101.903-24 – including *Taq* polymerase  
Lot number: 34M  
Expiry date: 2013-November-01  
Number of tests: 24  
Number of wells per test: 15 + 1  
Storage - pre-aliquoted primers: dark at -20°C  
- PCR Master Mix: -20°C  
- Adhesive PCR seals RT  
- Product Insert RT

**This Product Description is only valid for Lot No. 34M.**

### **CHANGES COMPARED TO THE PREVIOUS *OLERUP SSP*<sup>®</sup> DQA1\*02,05;DQB1\*02,03:02 Lot**

The specificity and interpretation tables have been updated for the DQA1 and DQB1 alleles described since the previous *Olerup SSP*<sup>®</sup> DQA1\*02,05;DQB1\*02,03:02 lot (**Lot No. 50K**) was made.

The primers of the wells detailed below have been exchanged, added or modified compared to the previous lot.

Well	5'-primer	3'-primer	rationale
8	Added	-	5'-primer added for the DQB1*03:37 allele.
9	Added	Added	Primer pair added for the DQB1*03:32 allele.

Change in revision R01 compared to R00:

1. The product length of the control primer pair in the Negative Control primer mix has been corrected in the specificity and interpretation tables.

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Well **16** contains Negative Control primer pairs, that will amplify more than 95% of the *Olerup SSP*<sup>®</sup> HLA Class I, DRB, DQB1 and DPB1 amplicons as well as the amplicons generated by control primer pairs.

PCR product sizes range from 75 to 430 base pairs.  
 The PCR product generated by the control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80
<b>5'-primer<sup>1</sup></b>	<b>164</b>	<b>340</b>	<b>440</b>	<b>45</b>	<b>45</b>	<b>43</b>
	5'-CAC <sup>3'</sup>	5'-Agg <sup>3'</sup>	5'-TTA <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-Tgg <sup>3'</sup>	5'-Tgg <sup>3'</sup>
<b>3'-primer<sup>2</sup></b>	<b>231</b>	<b>2<sup>nd</sup> I</b>	<b>507</b>	<b>59</b>	<b>58</b>	<b>57</b>
	5'-TgC <sup>3'</sup>	5'-AAA <sup>3'</sup>	5'-TTg <sup>3'</sup>	5'-CTC <sup>3'</sup>	5'-ggC <sup>3'</sup>	5'-CTC <sup>3'</sup>
<b>A*</b>	<b>+</b>	<b>+</b>	<b>+</b>			
<b>B*</b>	<b>+</b>	<b>+</b>	<b>+</b>			
<b>C*</b>	<b>+</b>	<b>+</b>	<b>+</b>			
<b>DRB1</b>				<b>+</b>	<b>+</b>	
<b>DRB3</b>				<b>+</b>	<b>+</b>	
<b>DRB5</b>				<b>+</b>		
<b>DQB1</b>					<b>+</b>	
<b>DPB1</b>						<b>+</b>

<sup>1</sup>The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>2</sup>The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon or the 2<sup>nd</sup> intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

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## PRODUCT DESCRIPTION

### DQA1\*02,05;DQB1\*02,03:02 SSP subtyping

#### CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQA1\*02, DQA1\*05, DQB1\*02 and DQB1\*03:02 alleles.

*Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.*

#### PLATE LAYOUT

Each test consists of 16 PCR reactions in a 16 well PCR plate.

1 DQA1	2 DQA1	3 DQB1	4 DQB1	5 DQB1	6 DQB1	7 DQB1	8 DQB1
9 DQB1	10 DQB1	11 DQB1	12 DQA1	13 DQA1	14 DQA1	15 DQA1	16 Neg ctrl

The 16 well cut PCR plate is marked with '34M' in silver/gray ink.

Well No. 1 is marked with the Lot No '34M'.

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

Wells 1, 2, 12 to 15: DQA1 primers.

Wells 3 to 11: DQB1 primers.

Well 16: Negative control primers.

The PCR plates are covered with a PCR-compatible foil.

**Please note:** When removing each 16 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

#### UNIQUELY IDENTIFIED ALLELES

All the DQA1\*02 and DQA1\*05 alleles as well as all the DQB1\*02 and DQB1\*03 alleles, i.e. **DQA1\*02:01, DQA1\*05:01 to 05:11, DQB1\*02:01 to 02:06 and DQB1\*03:01 to 03:37**, recognized by the HLA Nomenclature Committee in April 2011<sup>1</sup> have been considered in the specificity and interpretation tables of the DQA1\*02,05;DQB1\*02,03:02 kit.

<sup>1</sup>DQA1 and DQB1 alleles listed on the IMGT/HLA web page 2011-April-08, release 3.4.0, [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla).

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## SPECIFICITY TABLE

### DQA1\*02,05;DQB1\*02,03:02 SSP subtyping

Specificities and sizes of the PCR products of the 15+1 primer mixes used for DQA1\*02,05;DQB1\*02,03:02 SSP typing

Primer Mix	Size of spec. PCR product <sup>1</sup>	Size of control band <sup>2</sup>	Amplified DQA1 alleles <sup>3</sup>	Amplified DQB1 alleles <sup>3</sup>
<b>1</b>	175 bp	<b>430 bp</b>	*02:01	
<b>2</b>	165 bp	515 bp	*05:01:01:01-05:09, 05:11	
<b>3<sup>6</sup></b>	210 bp	515 bp		*02:01:01-02:06
<b>4<sup>4,5</sup></b>	80 bp	515 bp		*03:01:01:01-03:01:06, 03:03:02:01-03:03:03, 03:09-03:10, 03:12-03:13, 03:15-03:17, 03:19-03:24, 03:26-03:31, 03:33-03:36, *06:01:01 <sup>w</sup> -06:01:06 <sup>w</sup> , 06:07:01 <sup>w</sup> -06:07:02 <sup>w</sup> , 06:15 <sup>w</sup> , 06:32 <sup>w</sup> , 06:35 <sup>w</sup> , 06:37 <sup>w</sup>
<b>5<sup>5,6</sup></b>	135 bp	<b>430 bp</b>		*02:01:01-02:02, 02:04-02:06, *03:02:01-03:02:05, 03:07-03:08, 03:11, 03:18, 03:32, 03:37, *06:29
<b>6</b>	220 bp	515 bp		*02:01:01-02:06, *03:02:01-03:03:03, 03:06-03:08, 03:11-03:12, 03:15, 03:18, 03:20, 03:23, 03:25-03:26, 03:30-03:34, 03:37, *04:03:01-04:03:02
<b>7<sup>4,6</sup></b>	115 bp	515 bp		*03:06, 03:25
<b>8<sup>6,7</sup></b>	150 bp, 190 bp, 220 bp	515 bp		*03:07, 03:18, 03:37
<b>9<sup>8</sup></b>	135 bp, 175 bp	515 bp		*03:08, 03:32, *06:02:02, 06:03:02
<b>10<sup>9</sup></b>	135 bp, 260 bp	515 bp		*03:09, 03:11, 03:26
<b>11<sup>6,10</sup></b>	145 bp, 185 bp	515 bp		*03:01:01:01-03:37
<b>12<sup>4</sup></b>	90 bp	515 bp	*05:02	
<b>13</b>	200 bp	<b>430 bp</b>	*05:01:01:01-05:01:02, 05:02 <sup>?</sup> , 05:04 <sup>?</sup> , 05:05:01:01-05:05:01:03, 05:08-05:11	
<b>14</b>	200 bp	515 bp	*05:02 <sup>?</sup> , 05:03, 05:04 <sup>?</sup> , 05:06-05:07	

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<b>15</b>	205 bp	515 bp	*05:01:01:01- 05:03, 05:05:01:01- 05:09, 05:11
<b>16<sup>11</sup></b>	-	-	<b>Negative control</b>

<sup>1</sup> Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQA1 and DQB1 SSP typings.

When the primers in a primer mix can give rise to specific PCR products of more than one length this is indicated if the size difference is 20 base pairs or more. Size differences shorter than 20 base pairs are not given. For high resolution SSP kits the respective lengths of the specific PCR product(s) of the alleles amplified by these primer mixes are given.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

<sup>2</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 515 base pairs, for most wells, or a band of 430 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the shorter, 430 bp, internal positive control band in order to help in the correct orientation of the DQA1\*02,05;DQB1\*02,03:02 typing.

In addition, wells number 5 and 13 contain the primer pair giving rise to the shorter, 430 bp, internal positive control band in order to allow kit identification.

In the presence of a specific amplification the intensity of the control band often decreases.

<sup>3</sup>For several DQA1 and DQB1 alleles only partial second exon nucleotide sequences are available or nucleotide sequence information is not available for the 1<sup>st</sup> and 3<sup>rd</sup> exons. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of DQA1 and DQB1 alleles are conserved within allelic groups.

<sup>4</sup>Specific PCR fragments shorter than 125 base pairs are less intense and not as sharp as longer specific bands.

<sup>5</sup>Primer mixes 4 and 5 may yield less specific PCR products than the other DQB1 primer mixes.

<sup>6</sup>Primer mixes 3, 5, 7, 8 and 11 have a tendency of primer oligomer formation.

<sup>7</sup>Primer mix 8: Specific PCR fragment of 150 bp in the DQB1\*03:07 allele. Specific PCR fragment of 190 bp in the DQB1\*03:37 allele. Specific PCR fragment of 220 bp in the DQB1\*03:18 allele.

<sup>8</sup>Primer mix 9: Specific PCR fragment of 135 bp in the DQB1\*03:08 and the DQB1\*06:02:02 and 06:03:02 alleles. Specific PCR fragment of 175 bp in the DQB1\*03:32 alleles.

<sup>9</sup>Primer mix 10: Specific PCR fragment of 135 bp in the DQB1\*03:09 allele. Specific PCR fragment of 260 bp in the DQB1\*03:11 and 03:26 alleles.

<sup>10</sup>Primer mix 11: Specific PCR fragment of 145 bp in the DQB1\*03:01:02, 03:02:03, 03:03:03, 03:05:01-03:05:02, 03:20, 03:23 and 03:37 alleles. Specific PCR fragment of 185 bp in the DQB1\*03:25 allele. Specific PCR fragment of 145 and 185 bp in the DQB1\*03:01:01:01-03:01:01:03, 03:01:03-03:02:02, 03:02:04-03:03:02:03, 03:04, 03:05:03-03:19, 03:21-03:22, 03:24 and 03:26-03:36 alleles. Both bands may not always be visible.

<sup>11</sup>Primer mix 16 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by control primer pairs. PCR product sizes range from 75 to 200 base pairs. The PCR product generated by the control primer pair is 430 base pairs.

‘w’, might be weakly amplified.

‘?’, nucleotide sequence information not available for the primer matching sequence.

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INTERPRETATION TABLE								
DQA1*02,05;DQB1*02,03:02 typing								
Amplification patterns of the DQA1*02 and 05 and DQB1*02 and 03 alleles								
	Well <sup>5</sup>							
	1	2	3	4	5	6	7	8
Length of spec. PCR product(s)	175	165	210	80	135	220	115	150
								190
								220
Length of int. pos. control <sup>1</sup>	430	515	515	515	430	515	515	515
5'-primer(s) <sup>2</sup>	7 (90)	34 (169)	30 (185)	57 (266)	26 (173)	26 (173)	38 (210)	27 (175)
	5'-CAC <sup>3'</sup>	5'-AgC <sup>3'</sup>	5'-AAg <sup>3'</sup>	5'-TgA <sup>3'</sup>	5'-TCT <sup>3'</sup>	5'-TCT <sup>3'</sup>	5'-gCA <sup>3'</sup>	5'-TTC <sup>3'</sup>
								36 (204)
								5'-gAC <sup>3'</sup>
								49 (242)
								5'-ggT <sup>3'</sup>
3'-primer(s) <sup>3</sup>	52 (224)	75 (293)	86 (353)	70 (304)	57 (266)	86 (353)	62 (282)	86 (353)
	5'-TgT <sup>3'</sup>	5'-gAC <sup>3'</sup>	5'-gCT <sup>3'</sup>	5'-CCT <sup>3'</sup>	5'-Cgg <sup>3'</sup>	5'-gCT <sup>3'</sup>	5'-CTA <sup>3'</sup>	5'-gCT <sup>3'</sup>
Well No.	1	2	3	4	5	6	7	8
DQA1 or DQB1 allele <sup>4</sup>								
DQA1*02:01	1							
DQA1*05:01:01:01-05:01:02, 05:05:01:01-05:05:01:03, 05:08-05:09, 05:11		2						
DQA1*05:02		2						
Well No.	1	2	3	4	5	6	7	8

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INTERPRETATION TABLE								
DQA1*02,05;DQB1*02,03:02 typing								
Amplification patterns of the DQA1*02 and 05 and DQB1*02 and 03 alleles								
Well <sup>5</sup>								
9	10	11	12	13	14	15	16	
135	135	145	90	200	200	205	Neg. Ctrl.	
175	260	185						
515	515	515	515	430	515	515		
Length of spec. PCR product(s)								
Length of int. pos. control <sup>1</sup>								
38	14	38	59	107	107	21	Negative Control	
(210)	(136)	(210)	(245)	(389)	(389)	(131)		5'-primer(s) <sup>2</sup>
5'-gCA <sup>3'</sup>	5'-gCC <sup>3'</sup>	5'-gCA <sup>3'</sup>	5'-CCg <sup>3'</sup>	5'-CAT <sup>3'</sup>	5'-CAT <sup>3'</sup>	5'-TCC <sup>3'</sup>		
141	135	48						
(517)	(500)	(240)						
5'-CTA <sup>3'</sup>	5'-TgA <sup>3'</sup>	5'-CgC <sup>3'</sup>						
		55						
		(260)						
		55						
		(260)						
		5'-gCA <sup>3'</sup>						
70	86	86	75	160	160	75	3'-primer(s) <sup>3</sup>	
(304)	(353)	(353)	(293)	(547)	(547)	(293)		
5'-CCC <sup>3'</sup>	5'-gCT <sup>3'</sup>	5'-gCT <sup>3'</sup>	5'-gAC <sup>3'</sup>	5'-AgC <sup>3'</sup>	5'-AgA <sup>3'</sup>	5'-gAC <sup>3'</sup>		
185	167							
(650)	(596)							
5'-CgA <sup>3'</sup>	5'-CAT <sup>3'</sup>							
9	10	11	12	13	14	15	16	
Well No.								
DQA1 or DQB1 allele <sup>4</sup>								
DQA1*02:01								
DQA1*05:01:01:01-05:01:02, 05:05:01:01-05:05:01:03, 05:08-05:09, 05:11								
DQA1*05:02								
9	10	11	12	13	14	15	16	
Well No.								



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Length of spec.	175	165	210	80	135	220	115	150
PCR product(s)								190
								220
Well No.	1	2	3	4	5	6	7	8
DQA1 or DQB1 allele <sup>4</sup>								
DQA1*05:03, 05:06-05:07		2						
DQA1*05:04		2						
DQA1*05:10								
DQB1*02:01:01-02:02, 02:04-02:06			3		5	6		
DQB1*02:03			3			6		
DQB1*03:01:01:01-03:01:06, 03:10, 03:13, 03:16-03:17, 03:19, 03:21-03:22, 03:24, 03:27-03:29, 03:35-03:36				4				
DQB1*03:02:01-03:02:05					5	6		
DQB1*03:03:02:01-03:03:03, 03:12, 03:15, 03:20, 03:23, 03:30-03:31, 03:33-03:34				4		6		
DQB1*03:04-03:05:04, 03:14								
DQB1*03:06, 03:25						6	7	
DQB1*03:07, 03:18, 03:37					5	6		8
DQB1*03:08, 03:32					5	6		
DQB1*03:09				4				
DQB1*03:11					5	6		
DQB1*03:26				4		6		
DQB1*04:03:01-04:03:02						6		
DQB1*06:01:01-06:01:06, 06:07:01- 06:07:02, 06:15, 06:32, 06:35, 06:37				w				
DQB1*06:02:02, 06:03:02								
DQB1*06:29					5			
DQA1 or DQB1 allele <sup>4</sup>								
Well No.	1	2	3	4	5	6	7	8

<sup>1</sup>The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 515 base pairs, for most wells, or a band of 430 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the shorter, 430 bp, internal positive control band in order to help in the correct orientation of the DQA1\*02,05;DQB1\*02,03:02 typing.

In addition, wells number 5 and 13 contain the primer pair giving rise to the shorter, 430 bp, internal positive control band in order to allow kit identification.

<sup>2</sup>The codon, and in parenthesis the nucleotide, in the 1<sup>st</sup>, 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given. Codon and nucleotide numbering as in [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla). The sequence of the 3 terminal nucleotides of the primer is given.

<sup>3</sup>The codon, and in parenthesis the nucleotide, in the 2<sup>nd</sup> or 3<sup>rd</sup> exon, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Codon and nucleotide numbering as on the [www.ebi.ac.uk/imgt/hla](http://www.ebi.ac.uk/imgt/hla) web site. The sequence of the 3 terminal nucleotides of the primer is given.

<sup>4</sup>DQA1\*05013 has been renamed to DQA1\*05:05.

The sequence of the DQB1\*03031 allele has been shown to be identical to DQB1\*03:03:02.



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135	135	145	90	200	200	205			Length of spec. PCR product(s)
175	260	185							
9	10	11	12	13	14	15	16		Well No.
									DQA1 or DQB1 allele <sup>4</sup>
					14	15			DQA1*05:03, 05:06-05:07
				?	?				DQA1*05:04
				13					DQA1*05:10
									DQB1*02:01:01-02:02, 02:04-02:06
									DQB1*02:03
		11							DQB1*03:01:01:01-03:01:06, 03:10, 03:13, 03:16-03:17, 03:19, 03:21-03:22, 03:24, 03:27-03:29, 03:35-03:36
		11							DQB1*03:02:01-03:02:05
		11							DQB1*03:03:02:01-03:03:03, 03:12, 03:15, 03:20, 03:23, 03:30-03:31, 03:33-03:34
		11							DQB1*03:04-03:05:04, 03:14
		11							DQB1*03:06, 03:25
		11							DQB1*03:07, 03:18, 03:37
9		11							DQB1*03:08, 03:32
	10	11							DQB1*03:09
	10	11							DQB1*03:11
	10	11							DQB1*03:26
									DQB1*04:03:01-04:03:02
									DQB1*06:01:01-06:01:06, 06:07:01- 06:07:02, 06:15, 06:32, 06:35, 06:37
9									DQB1*06:02:02, 06:03:02
									DQB1*06:29
									DQA1 or DQB1 allele <sup>4</sup>
9	10	11	12	13	14	15	16		Well No.

Negative Control

<sup>5</sup>Primer mix 8: Specific PCR fragment of 150 bp in the DQB1\*03:07 allele. Specific PCR fragment of 190 bp in the DQB1\*03:37 allele. Specific PCR fragment of 220 bp in the DQB1\*03:18 allele.

Primer mix 9: Specific PCR fragment of 135 bp in the DQB1\*03:08 and the DQB1\*06:02:02 and 06:03:02 alleles. Specific PCR fragment of 175 bp in the DQB1\*03:32 alleles.

Primer mix 10: Specific PCR fragment of 135 bp in the DQB1\*03:09 allele. Specific PCR fragment of 260 bp in the DQB1\*03:11 and 03:26 alleles.

Primer mix 11: Specific PCR fragment of 145 bp in the DQB1\*03:01:02, 03:02:03, 03:03:03, 03:05:01-03:05:02, 03:20, 03:23 and 03:37 alleles. Specific PCR fragment of 185 bp in the DQB1\*03:25 allele. Specific PCR fragment of 145 and 185 bp in the DQB1\*03:01:01:01-03:01:01:03, 03:01:03-03:02:02, 03:02:04-03:03:02:03, 03:04, 03:05:03-03:19, 03:21-03:22, 03:24 and 03:26-03:36 alleles. Both bands may not always be visible.

Primer mix 16 contains a negative control, which will amplify more than 95% of HLA amplicons as well as the amplicons generated by control primer pairs. PCR product sizes range from 75 to 200 base pairs. The PCR product generated by the control primer pair is 430 base pairs.

‘w’, might be weakly amplified.

‘?’ , nucleotide sequence information not available for the primer matching sequence.

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<b>CELL LINE VALIDATION SHEET</b>									
<b>DQA1*02,05;DQB1*02,03:02 typing kit</b>				<b>Well</b>					
				<b>1</b>	<b>2</b>	<b>12</b>	<b>13</b>	<b>14</b>	<b>15</b>
			Prod. No.:	201076701	201076702	201076713	201076714	201076715	201076716
	<b>IHWC cell line</b>	<b>DQA1*</b>							
1	9001 SA	*01:01		-	-	-	-	-	-
2	9280 LK707	*01:03	*03:03	-	-	-	-	-	-
3	9011 E4181324	*01:03		-	-	-	-	-	-
4	9275 GU373	*05:01		-	+	-	+	-	+
5	9009 KAS011	*01:02		-	-	-	-	-	-
6	9353 SM	*01:03	*03:01	-	-	-	-	-	-
7	9020 QBL	*05:01		-	+	-	+	-	+
8	9025 DEU	*03		-	-	-	-	-	-
9	9026 YAR	*03:01		-	-	-	-	-	-
10	9107 LKT3	*03:03		-	-	-	-	-	-
11	9051 PITOUT	*02:01		+	-	-	-	-	-
12	9052 DBB	*02:01		+	-	-	-	-	-
13	9004 JESTHOM	*01:01		-	-	-	-	-	-
14	9071 OLGA	*04:01		-	-	-	-	-	-
15	9075 DKB	*03:02		-	-	-	-	-	-
16	9037 SWEIG007	*05:05		-	+	-	+	-	+
17	9282 CTM3953540	*01:03	*05:01	-	+	-	+	-	+
18	9257 32367	*01:02	*03:03	-	-	-	-	-	-
19	9038 BM16	*05:05		-	+	-	+	-	+
20	9059 SLE005	*01:02		-	-	-	-	-	-
21	9064 AMALA	*05:03		-	+	-	-	+	+
22	9056 KOSE	*01:02	*01:04	-	-	-	-	-	-
23	9124 IHL	*01:03	*01:04	-	-	-	-	-	-
24	9035 JBUSH	*05:05		-	+	-	+	-	+
25	9049 IBW9	*02:01		+	-	-	-	-	-
26	9285 WT49	*05:01		-	+	-	+	-	+
27	9191 CH1007	*03:03	*01:05	-	-	-	-	-	-
28	9320 BEL5GB	*02:01	*03:03	+	-	-	-	-	-
29	9050 MOU	*02:01		+	-	-	-	-	-
30	9021 RSH	*04:01		-	-	-	-	-	-
31	9019 DUCAF	*05:01		-	+	-	+	-	+
32	9297 HAG	*05:05		-	+	-	+	-	+
33	9098 MT14B	*03:01		-	-	-	-	-	-
34	9104 DHIF	*05:05		-	+	-	+	-	+
35	9302 SSTO	*03:01		-	-	-	-	-	-
36	9024 KT17	*03:01		-	-	-	-	-	-
37	9065 HHKB	*01:03		-	-	-	-	-	-
38	9099 LZL	*05:03		-	+	-	-	+	+
39	9315 CML	*03:03	*05:01	-	+	-	+	-	+
40	9134 WHONP199	*02:01	*03:02	+	-	-	-	-	-
41	9055 H0301	*01:02		-	-	-	-	-	-
42	9066 TAB089	*01:03		-	-	-	-	-	-
43	9076 T7526	*03:02		-	-	-	-	-	-
44	9057 TEM	*01:04		-	-	-	-	-	-
45	9239 SHJO	*02:01	*03:03	+	-	-	-	-	-
46	9013 SCHU	*01:02		-	-	-	-	-	-
47	9045 TUBO	*05:05		-	+	-	+	-	+
48	9303 TER-ND	*01:01		-	-	-	-	-	-



Lot No.: **34M**

Lot-specific information

[www.olerup-ssp.com](http://www.olerup-ssp.com)

CELL LINE VALIDATION SHEET												
DQA1*02,05;DQB1*02,03:02 typing kit												
				Well								
				3	4	5	6	7	8	9	10	11
				201076703	201076704	201076705	201076706	201076707	201187608	201187609	201076710	201076711
IHWC cell line		DQB1		Production No.								
1	9001 SA	*05:01		-	-	-	-	-	-	-	-	-
2	9280 LK707	*06:01	*02:02	+	W	+	+	-	-	-	-	-
3	9011 E4181324	*06:01		-	W	-	-	-	-	-	-	-
4	9275 GU373	*02:01		+	-	+	+	-	-	-	-	-
5	9009 KAS011	*05:02		-	-	-	-	-	-	-	-	-
6	9353 SM	*03:02	*06:01	-	+	+	+	-	-	-	-	+
7	9020 QBL	*02:01		+	-	+	+	-	-	-	-	-
8	9025 DEU	*03:01		-	+	-	-	-	-	-	-	+
9	9026 YAR	*03:02		-	-	+	+	-	-	-	-	+
10	9107 LKT3	*04:01		-	-	-	-	-	-	-	-	-
11	9051 PITOUT	*02:02		+	-	+	+	-	-	-	-	-
12	9052 DBB	*03:03		-	+	-	+	-	-	-	-	+
13	9004 JESTHOM	*05:01		-	-	-	-	-	-	-	-	-
14	9071 OLGA	*04:02		-	-	-	-	-	-	-	-	-
15	9075 DKB	*03:03		-	+	-	+	-	-	-	-	+
16	9037 SWEIG007	*03:01		-	+	-	-	-	-	-	-	+
17	9282 CTM3953540	*02:01	*06:03	+	-	+	+	-	-	-	-	-
18	9257 32367	*06:02	*02:02	+	-	+	+	-	-	-	-	-
19	9038 BM16	*03:01		-	+	-	-	-	-	-	-	+
20	9059 SLE005	*06:04		-	-	-	-	-	-	-	-	-
21	9064 AMALA	*03:01		-	+	-	-	-	-	-	-	+
22	9056 KOSE	*05:03	*06:04	-	-	-	-	-	-	-	-	-
23	9124 IHL	*05:03	*06:01	-	W	-	-	-	-	-	-	-
24	9035 JBUSH	*03:01		-	+	-	-	-	-	-	-	+
25	9049 IBW9	*02:02		+	-	+	+	-	-	-	-	-
26	9285 WT49	*02:01		+	-	+	+	-	-	-	-	-
27	9191 CH1007	*04:01	*05:01	-	-	-	-	-	-	-	-	-
28	9320 BEL5GB	*02:02	*03:01	+	+	+	+	-	-	-	-	+
29	9050 MOU	*02:02		+	-	+	+	-	-	-	-	-
30	9021 RSH	*04:02		-	-	-	-	-	-	-	-	-
31	9019 DUCAF	*02:01		+	-	+	+	-	-	-	-	-
32	9297 HAG	*03:01		-	+	-	-	-	-	-	-	+
33	9098 MT14B	*03:02		-	-	+	+	-	-	-	-	+
34	9104 DHIF	*03:01		-	+	-	-	-	-	-	-	+
35	9302 SSTO	*03:05		-	-	-	-	-	-	-	-	+
36	9024 KT17	*03:02		-	-	+	+	-	-	-	-	+
37	9065 HHKB	*06:03		-	-	-	-	-	-	-	-	-
38	9099 LZL	*03:01		-	+	-	-	-	-	-	-	+
39	9315 CML	*02:01	*03:01	+	+	+	+	-	-	-	-	+
40	9134 WHONP199	*02:02	*03:03	+	+	+	+	-	-	-	-	+
41	9055 H0301	*06:09		-	-	-	-	-	-	-	-	-
42	9066 TAB089	*06:01		-	W	-	-	-	-	-	-	-
43	9076 T7526	*03:03		-	+	-	+	-	-	-	-	+
44	9057 TEM	*05:03		-	-	-	-	-	-	-	-	-
45	9239 SHJO	*02:02		+	-	+	+	-	-	-	-	-
46	9013 SCHU	*06:02		-	-	-	-	-	-	-	-	-
47	9045 TUBO	*03:01		-	+	-	-	-	-	-	-	+
48	9303 TER-ND	*05:01		-	-	-	-	-	-	-	-	-



Lot No.: **34M**

Lot-specific information

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## CERTIFICATE OF ANALYSIS

### **Olerup SSP<sup>®</sup> DQA1\*02,05;DQB1\*02,03:02 SSP**

**Product number:** 101.903-24 – including *Taq* polymerase  
**Lot number:** 34M  
**Expiry date:** 2013-November-01  
**Number of tests:** 24  
**Number of wells per test:** 15 + 1

#### **Well specifications:**

Well No.	Production No.	Well No.	Production No.
1	2010-767-01	9	2011-876-09
2	2010-767-02	10	2010-767-10
3	2010-767-03	11	2010-767-11
4	2010-767-04	12	2010-767-12
5	2010-767-05	13	2010-767-13
6	2010-767-06	14	2010-767-14
7	2010-767-07	15	2010-767-15
8	2011-876-08		

The specificity of each primer solution of the kit has been tested against 48 well characterized IHWC cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 7 to 10 and 12 were available. The specificities of the primers in primer solutions 7, 9 and 10 were tested by separately adding one additional 5'-primers, respectively one additional 3'-primer. In primer solutions 8 and 12 it was only possible to test the 3'-primers, the 5'-primers were not possible to test. In primer solution 9 one of the 5'-primers was not possible to test, and in primer solution 10 one of the 3'-primers was not possible to test.

The negative control primer pairs, **Production No. 2010-760-01**, can detect contamination with PCR products diluted  $10^{-7}$ .

**Results:** No false positive or false negative amplifications were obtained.

**Date of approval:** 2013-January-10

**Approved by:**

#### **Production Quality Control**

Lot No.: **34M**

Lot-specific information

[www.olerup-ssp.com](http://www.olerup-ssp.com)

## Declaration of Conformity

**Product name:** *Olerup* SSP® DQA1\*02,05;DQB1\*02,03:02  
**Product number:** 101.903-24  
**Lot number:** 34M

**Intended use:** DQA1\*02, DQA1\*05, DQB1\*02 and DQB1\*03 medium resolution histocompatibility testing

**Manufacturer:** *Olerup* SSP AB  
Franzengatan 5  
SE-112 51 Stockholm, Sweden  
**Phone:** +46-8-717 88 27  
**Fax:** +46-8-717 88 18

We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2008 and ISO 13485:2003, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex III, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Franzengatan 5, SE-112 51 Stockholm, Sweden.

Stockholm, Sweden  
2013-January-10

Ann-Cathrin Jareman  
Head of QA and Regulatory Affairs

Lot No.: **34M**

Lot-specific information

[www.olerup-ssp.com](http://www.olerup-ssp.com)

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**Web page:** <http://www.olerup.com>

For information on *Olerup* SSP distributors worldwide, contact **Olerup GmbH**.